Two New Compounds from Goniothalamus Cheliensis Hu

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Abstract: Two new compounds, gonioquinone (1) and goniofufurone acetonide (2), have been isolated from the roots of *Goniothalamus cheliensis* Hu. Their structures were determined on the basis of spectral and chemical evidence.

Keywords: Goniothalamus cheliensis Hu, gonioquinone, goniofufurone acetonide.

Goniothalamus cheliensis Hu (Annonaceace) is a small tree or shrub in Yunnan Province, southwest of China. Acetogenins, styryllactones and alkaloids¹⁻³ from the genus were reported in the previous literature and many of them showed strong cytotoxic activities against a number of human cancer cell lines⁴. The titled plant has been the subject of our investigation due to the impressive cytotoxicity of its organic extract against mouse lymphocytic leukemia cells in preliminary pharmacological screening. Bioassy-guided fractionation of the EtOH extract of the roots resulted in the isolation of two new compounds, together with thirty-six known compounds. In this paper, we report the structure elucidation of the new compounds (1 and 2).

Figure 1 Structure and key HMBC correlations for 1



Gonioquinone (1) was isolated as orange needles, mp 201-202 °C. Its molecular formula was determined as $C_{13}H_{10}O_5$ by HRFABMS at m/z 246.0907 [M]⁺ (calcd. 246.0892).

The IR spectrum (KBr, cm⁻¹) displayed absorption bands for hydroxyl (3379), carbonyl (1685) and aromatic moiety (1616 and 1589). Its UV spectrum exhibited characteristic bands at λ_{max} (loge): 410 (3.73), 285 (4.19), 234 (4.24), and 210 (4.29) nm analogous to those of draserone and suggested that **1** had a naphthaquinone skeleton⁵.

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The ¹H NMR spectrum of **1** showed the presence of two phenolic hydroxyl at δ 11.38 (s) and 10.07 (brs), three aromatic protons at δ 7.75(d, 1H, *J*=8.0Hz), 7.74 (t, 1H, *J*=8.0Hz) and 7.20 (d, 1H, *J*=8.0Hz) and the absence of signals in the quinonoid proton region (δ 5.8-6.8) indicating **1** to be a 1,4-naphthoquinone substituted at the 2- and 3- positions⁶. The ¹³C and DEPT NMR spectra displayed 13 carbon signals including one primary, one secondary, three tertiary and eight quarternary

Table 1 ¹H (500MHz) and ¹³C (125MHz) NMR data of 1 in CDCl₃

Position	$\delta_{\rm H} J$ (Hz)	δ_{C}	DEPT	Position	$\delta_{\rm H} J$ (Hz)	δ_{C}	DEPT
1		180.4	s	7	7.74 t (8.0)	139.1	d
2	10.71brs	152.2	s	8	7.75 d (8.0)	119.7	d
3		109.2	s	8a		133.6	s
4		184.7	s	9	2.73 s	33.2	t
4a		113.9	s	10		202.2	s
5	11.38 s	161.8	s	11	1.25 s	29.7	q
6	7.20 d (8.0)	122.2	d				

Table 2 1 H (500MHz) and 13 C(125MHz) NMR data of 2 in CDCL $\delta_{\rm H} J$ (Hz) $\delta_{\rm C}$ Position $\delta_{\rm H} J$ (Hz) $\delta_{\rm H} J$

Position	$\delta_{\rm H} J$ (Hz)	$\delta_{\rm C}$	Position	$\delta_{\rm H} J$ (Hz)	δ _C
2		174.5	10		101.8
3	2.71 d (3.0)	36.5	12	1.46 s	23.9
4	5.06 m	78.3	13	1.47 s	24.4
5	4.95 d (3.5)	87.0	1'		139.2
6	4.56 dd (8.0)	74.8	2'(6')	7.41 m	126.3
7	4.44 dd (4.5, 8.0)	84.9	3'(5')	7.36 m	128.5
8	4.65 d (4.5)	72.0	4'	7.31 m	128.0

carbons, which were identical with those of julomycin F except for the signal at δ 109.2 in the ¹³C NMR spectrum suggesting that **1** had the same skeleton and substituted groups as julomycin \vec{F} . A combination of the ¹H and ¹³C NMR spectra of **1** revealed the resonances for the protons at δ 2.73 (s, 2H) and 1.25 (s, 3H) and the carbons at δ 29.7, 202.2 and 33.2 indicating the presence of the fragment -CH₂COCH₃ (**Table 1**)⁸. The more shielded carbonyl resonance in the ¹³C NMR spectrum was attributable to C-1 (δ 180.4) because of the existence of a electron-repelling phenolic hydroxyl in the vicinity of C-1⁹.

Another hydroxyl was located at C-5 position, this finding was confirmed by the detailed analysis of J value and 2D NMR spectra. Three aromatic protons at δ 7.75, 7.74 and 7.20 with J values of 8.0Hz suggested that they were adjacent to each other. Combination with the HMQC spectrum, these protons were assigned to be connected with the carbons at δ 119.7 (C-8), 139.1 (C-7) and 122.2 (C-6) respectively. Furthermore, the HMBC correlations of the proton at δ_H 11.38 (OH) with the carbon signals at δ_C 161.8 (C-5) and 122.2 (C-6) revealed that the phenolic hydroxyl was attached to C-5 position.

The left acetonyl was linked to C-3 position, it was determined by the HMBC correlations of the proton at δ_H 2.73 (H-9) with the carbon signals at δ_C 109.2 (C-3) and 202.2 (C-10). In the ¹³C NMR spectrum, the signal for C-3 in **1** showed a more upfield

shift than C-3 of juglomycin F, it probably resulted from the formation of intramolecular hydrogen bond between the carbonyl group of C-4 and the phenolic hydroxyl proton of $C-5^7$.

Goniofufurone acetonide **Q**), mp 165-166°C, $[\alpha]_D^{23}$ +80 (c 0.05, MeOH), was obtained as white crystals. Its HRFABMS gave a quasi-molecular ion $[MH]^+$ at m/z 291.1248 (calcd. 291.1232), which fits the molecular formula $C_{16}H_{18}O_5$.

The IR spectrum (KBr, cm⁻¹) exhibited absorption bands for saturated γ -lactone (1786) and phenyl (1603 and 1495). Its ¹H NMR spectrum revealed the presence of five aromatic protons at δ 7.31-7.41 (m, 5H), five oxygenated methine protons at δ 4.44 -5.06, two methylene protons at δ 2.71 (d, 2H) and six methyl protons at δ 1.46 (s, 3H) and 1.47 (s, 3H) (**Table 2**), which were similar to those of goniofufurone (**3**) except for the signals of two methyl groups. By comparison of the ¹³C NMR data with those of

Figure 2 Structures for 2 and goniofufrone (3)



goniofufurone, **2** had an additional signal at δ 101.8 and two additional methyl signals at δ 23.9 and 24.4, which suggested that **2** was acetone derivative of goniofufurone. This was confirmed by the preparation of **2** from **3** reacting with 2, 2-dimethoxyl propane, the derivative showed the same data of ¹H NMR and R_f value as **2**. So **2** has the same absolute configurations as goniofufurone namely 4R, 5S, 6R, 7R, 8R¹⁰.

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References

- 1. X. P. Fang, A. Jone, C. J. Chang, J. L. Mclaughlin, J. Nat. Prod., 1991, 54, 1034.
- 2. S. Noppamas, S. Khanit, B. Rapepol, P. Chamnan, M. C. John, J. Nat. Prod., 1999, 62, 1390.
- 3. Z. Jiang, D. Q. Yu, J. Nat. Prod. 1997, 60, 122.
- 4. J. X. Zhu, J. G. Yu, Z. X. Luo, L. Sun, D. Y. Li, S. L. Yang, *Chinese Traditional and Herbal* Drugs, 2000, 31, 813.

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- 5. L. Huang, D.Q Yu, *The Application of UV Spectroscopy in Organic Chemistry*, No. 2, Science Press, **2000**, p.546.
- 6. K. Inoue, C. C. Chen, H. Inouye, K. Kuriayama, J. Chem. Soc. Perkin I, 1981, 2764.
- 7. H. Lessman, J. Krupa, H. Lacker, Z. Naturforcsh. B, 1989, 44, 353.
- 8. J. H. Tatum, R. A. Baker, R. E. Berry, *Phytochemistry*, **1985**, 24, 457.
- 9. G. K. Poch, J. B. Gloer, J. Nat. Prod., 1992, 55, 1093.
- 10. K. M. T. Shing, H. C. Tsui, J. Chem. Soc. commun., 1992. 432.

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